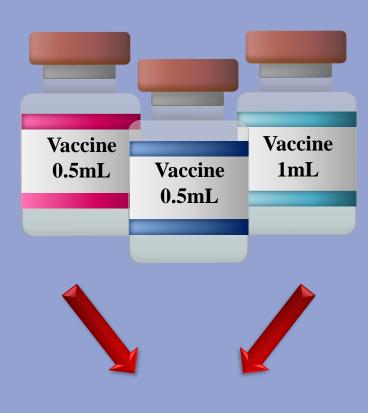
# Exploring the rationale for the high concentration of aluminium used in clinical vaccinations.







### Introduction



How does the concentration of AI in vaccines impact the biology at the site of injection?

### Maximum amount of AI permitted in vaccines [1]

- ➤ 0.85mg/dose if determined by assay
- ➤ 1.14mg/dose if determined by calculation on basis of Al compound added

#### **Average amount of AI in vaccines**

ca 0.4mg/dose (0.8mg/mL)

#### Rationale

- ➤ Higher concentrations of Al are more effective.
  - ❖>0.5mg/dose Al did not improve efficacy of tetanus vaccine [2].
  - ❖ High concentrations of AI may impede immunological response – cytotoxicity [3]

### Methodological approach



#### **Vaccine models**

➤ Alhydrogel in saline (no antigen) – 0.3-0.9mg/mL Al (pH 7)

#### **Physicochemical characterisation**

- Particle size
- > Zeta potential
- ➤ Size exclusion filtration/GFAAS (Al quantification)

#### **Uptake**

- ➤ Fluorescence microscopy lumogallion staining (50µM)
- ➤ Macrophages exposed to Al for 1hr

#### **Cell viability**

Presto blue assay – determinant of metabolic activity (resazurin based assay)

## DLS – Particle size distribution vs. concentration of Al in simulated vaccines.

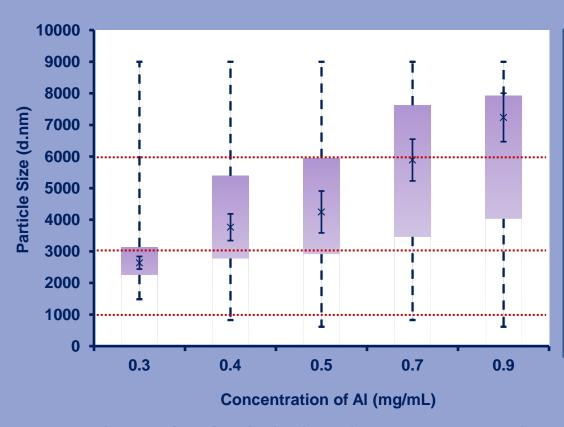


Fig 1: Particle size distributions of Alhydrogel only vaccines containing 0.3-0.9mg/mL Al. Purple boxes indicate the interquartile range of the data while the dashed bars show the span. Blue crosses show the average d50 values and d50 error bars represent the ±SD of the measurement where n=5. The red dotted lines highlight the relevant filtration size cut-offs used in complementary experiments.

- Interquartile range encompassed larger particles as the concentration of Al was increased.
- The breadth of the interquartile range also increased when the concentration of AI was increased.
- Based on the theoretical filtration size cut-offs:
  - ❖ The majority of the AI in these vaccines will exist as micron-sized aggregates i.e. >1µm.
  - ❖ Significant shifts in size are expected between 0.3 & 0.4mg/mL and 0.5 & 0.7mg/mL.

### DLS – PDI vs. concentration of AI in simulated vaccines.

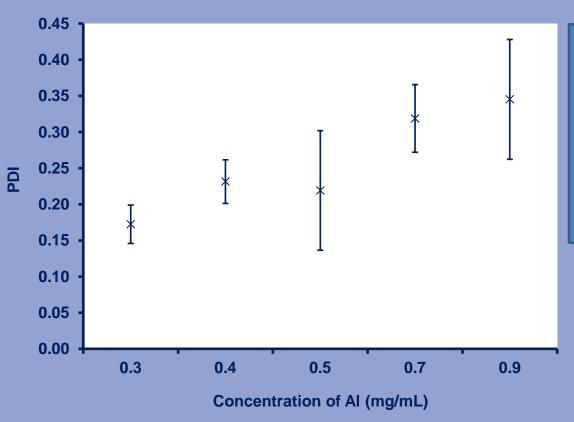


Fig 2: PDI of Alhydrogel only vaccines containing 0.3-0.9mg/mL Al. Error bars represent the ±SD of the measurement where n=5

- PDI values predominantly increased over the concentration range studied (0.172-0.345).
- Significant difference in PDI observed between :
  - **❖** 0.3 & 0.9mg/mL (0.172 vs. 0.345, P=0.001).
  - ❖ 0.4 & 0.7 mg/mL AI (0.231 vs. 0.319, P=0.05).

## DLS – Median particle size vs. concentration of Al in simulated vaccines.

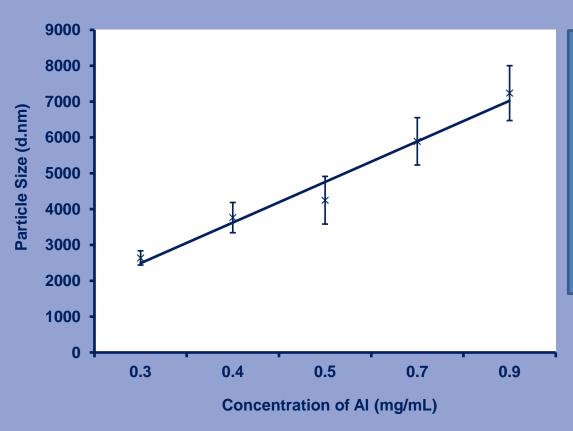


Fig 3: Particle size (d50) of Alhydrogel only vaccines containing 0.3-0.9mg/mL Al. Error bars represent the ±SD of the measurement where n=5

- D50 values increased in a linear manner over the concentration range studied (2638 -7237 nm, R<sup>2</sup> = 0.974).
- Significant difference in median particle size between:
  - ❖ 0.3 & 0.9 mg/mL (2638 nm vs. 7237 nm, P<0.0001).</p>
  - ❖ 0.3 & 0.4 mg/mL (2638 nm vs. 3764 nm, P=0.03)
  - ❖ 0.5 & 0.7 mg/mL (4247 nm vs. 5890 nm, P=0.0007).

## ELS – Zeta potential vs. concentration of Al in simulated vaccines.

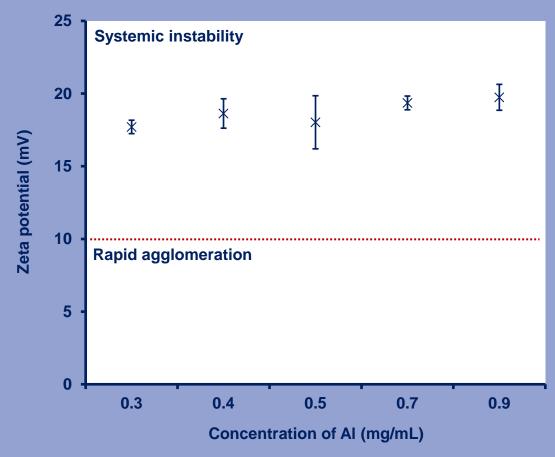
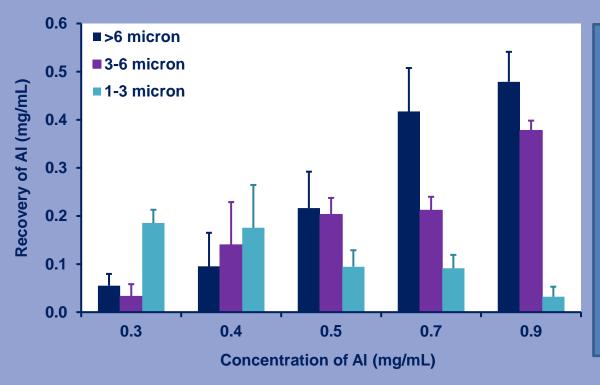


Fig 4: Zeta potential of Alhydrogel only vaccines containing 0.3-0.9mg/mL Al. Error bars represent the ±SD of the measurement where n=5

- Zeta potential values remained fairly consistent over the concentration range studied and were located in the region associated with systemic instability (17.71-19.74mV).
- Lowest vaccine dose had a significantly lower zeta potential than that of the highest vaccine dose (17.71 vs. 19.74 mV, P=0.05).
- Vaccine particles are positively charged at pH 7

## GFAAS/filtration – % aluminium recovery vs. concentration of AI in simulated vaccines.



Abundance of particles between 1-
3µm decreased when concentration
of Al increased.

Significant differences observed
between:

- 0.3mg/mL & 0.9mg/mL (0.19mg/mL vs. 0.03mg/mL, P=0.004)
- ❖ 0.4mg/mL & 0.9mg/mL
- (0.18mg/mL vs. 0.03mg/mL, P=0.01)
- ➤ Smaller particles present at 0.3mg/mL vs. 0.9mg/mL lower concentrations are more likely to be internalised by macrophages

Concentration of AI (mg/mL)	0.3	0.4	0.5	0.7	0.9
	Recovery of AI (%)				
1-3µm	77.06	55.04	10.89	3.45	0.68
3-6µm	2.71	17.30	47.02	33.64	30.09
>6µm	20.19	27.62	42.08	62.90	69.20

Fig 5: The relative abundance of aluminium (%) within specific particle size fractions for Alhydrogel only vaccines containing an initial concentration of 0.3-0.9mg/mL Al. Blue, purple and green boxes represent the % Al in the size fractions >6μm, 3-6μm & 1-3μm respectively.

## DLS – Particle size distribution of simulated vaccines in R10 medium (1hr incubation)

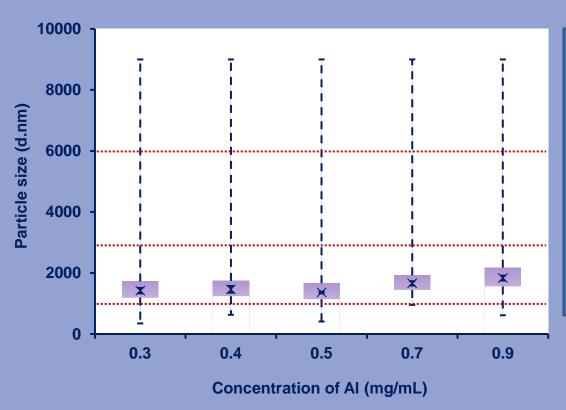


Fig 6: Particle size distributions of Alhydrogel only vaccines 0.3-0.9mg/mL Al following 1hr incubation within R10 medium (37°C). Purple boxes indicate the interquartile range of the data while the dashed bars show the span. Blue crosses show the average d50 values and d50 error bars represent the ±SD of the measurement where n=5. The red dotted lines highlight the relevant filtration size cut-offs used in complementary experiments.

- Interquartile range of the data remained consistent as the concentration of Al was increased.
- The breadth of the interquartile range also remained stable as the concentration of AI was increased.
- Based on the theoretical filtration size cut-offs:
  - ❖ The majority of the Al will exist as micron-sized aggregates i.e. >1µm.
  - Significant shifts in size between concentrations are unlikely.

# DLS – PDI of simulated vaccines in R10 medium (1hr incubation)

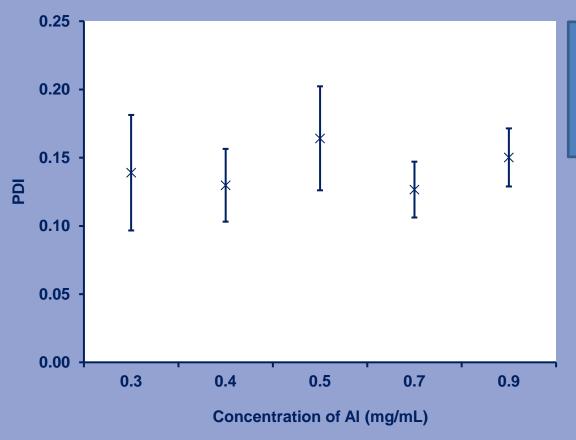


Fig 7: PDI of Alhydrogel only vaccines containing 0.3-0.9mg/mL Al following 1hr incubation within R10 medium (37°C). Error bars represent the ±SD of the measurement where n=5

- PDI values remained stable over the concentration range studied (0.127-0.164).
- No significant difference in PDI between any of the concentrations studied.

## DLS – Median particle size vs. concentration of Al in simulated vaccines.

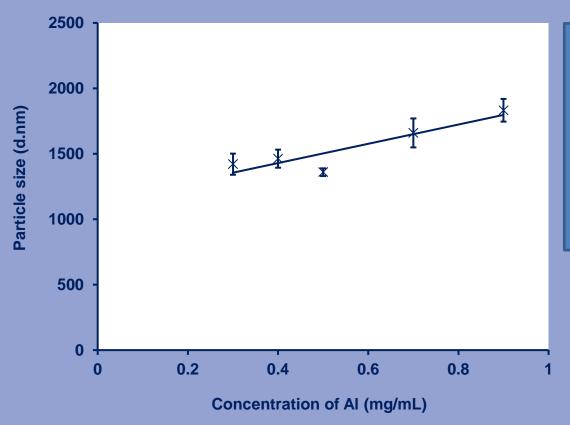


Fig 8: Particle size (d50) of Alhydrogel only vaccines containing 0.3-0.9mg/mL Al following 1hr incubation within R10 medium (37°C). Error bars represent the ±SD of the measurement where n=5

- ▶ D50 values increased in a weakly linear manner over the concentration range studied (2638 -7237nm, R² = 0.822).
- Significant difference in particle size between:
  - ❖ 0.3 & 0.9 mg/mL (1421 nm vs. 1833 nm, P<0.0001).
  - ❖ 0.5 & 0.7 mg/mL (1360 nm vs. 1660 nm, P<0.0001).</p>

## ELS – Zeta potential vs. concentration of Al in simulated vaccines.

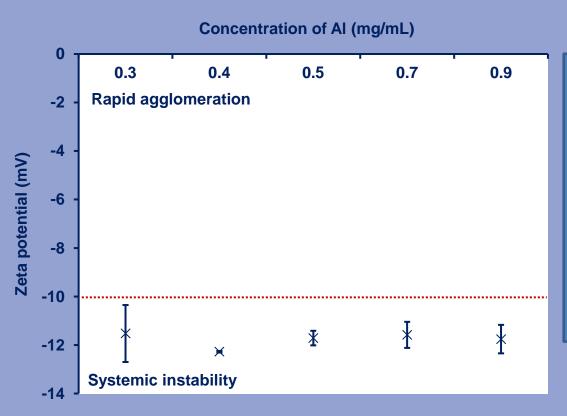
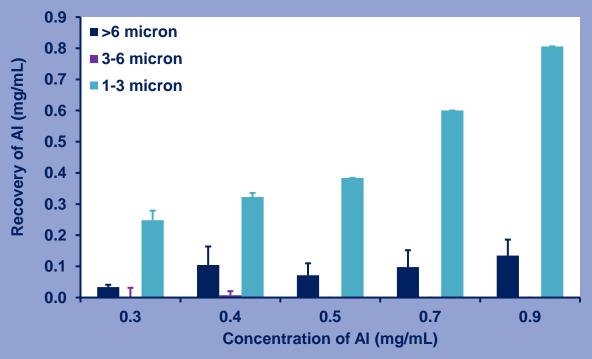


Fig 9: Zeta potential of Alhydrogel only vaccines containing 0.3-0.9mg/mL Al following 1hr incubation within R10 medium (37°C). Error bars represent the ±SD of the measurement where n=5

- Zeta potential values remained fairly consistent over the concentration range studied and were located in the region associated with systemic instability (-11.52- -12.28mV).
- No significant difference between concentrations of AI
- Vaccine particles are negatively charged when administered into R10 medium.
- Evidence of protein adsorption (surface saturation) at all concentrations studied.

# GFAAS/filtration – % aluminium recovery of simulated vaccines in R10 medium (1hr incubation)

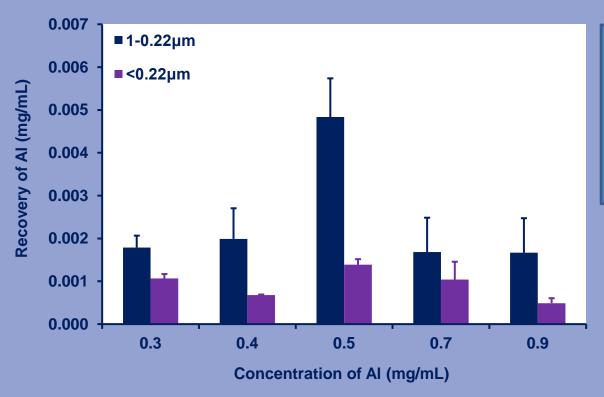


Concentration of AI (mg/mL)	0.3	0.4	0.5	0.7	0.9	
	Recovery of AI (%)					
1-3µm	87.43	73.56	83.46	85.99	85.69	
3-6µm	0.18	1.69	0	0	0	
>6µm	11.78	23.80	15.48	14.01	14.31	

- Abundance of particles between 1-3µm increased when concentration of Al increased.
- Significant differences observed between:
  - 0.3mg/mL & 0.7mg/mL(0.25mg/mL vs. 0.6 mg/mL, P=0.01)
  - 0.3mg/mL & 0.9mg/mL(0.25mg/mL vs. 0.8 mg/mL,P=0.0001)
  - ❖ 0.4 mg/mL & 0.9 mg/mL (0.32 mg/mL vs. 0.8mg/mL, P=0.01
- Larger availability of smaller particles at higher concentrations of Al

Fig 10: The relative abundance of aluminium (%) within specific particle size fractions for Alhydrogel only vaccines containing an initial concentration of 0.3-0.9mg/mL Al following 1 hr incubation within R10 medium (37°C). Blue, purple and green boxes represent the % Al in the size fractions >6µm, 3-6µm & 1-3µm respectively.

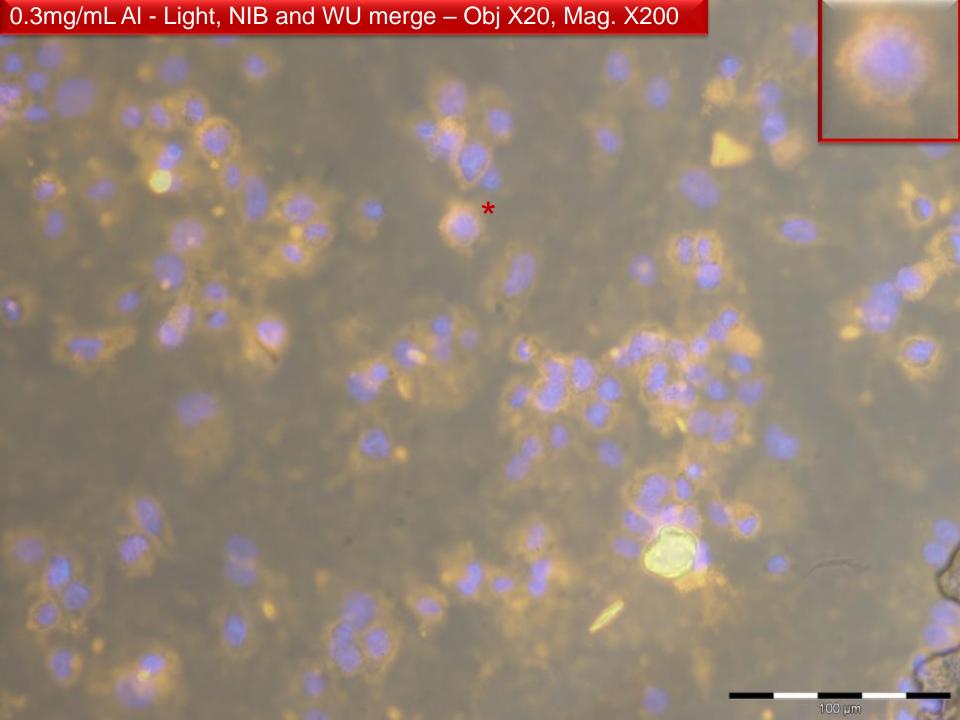
# GFAAS/filtration – % aluminium recovery of simulated vaccines in R10 medium (1hr incubation)

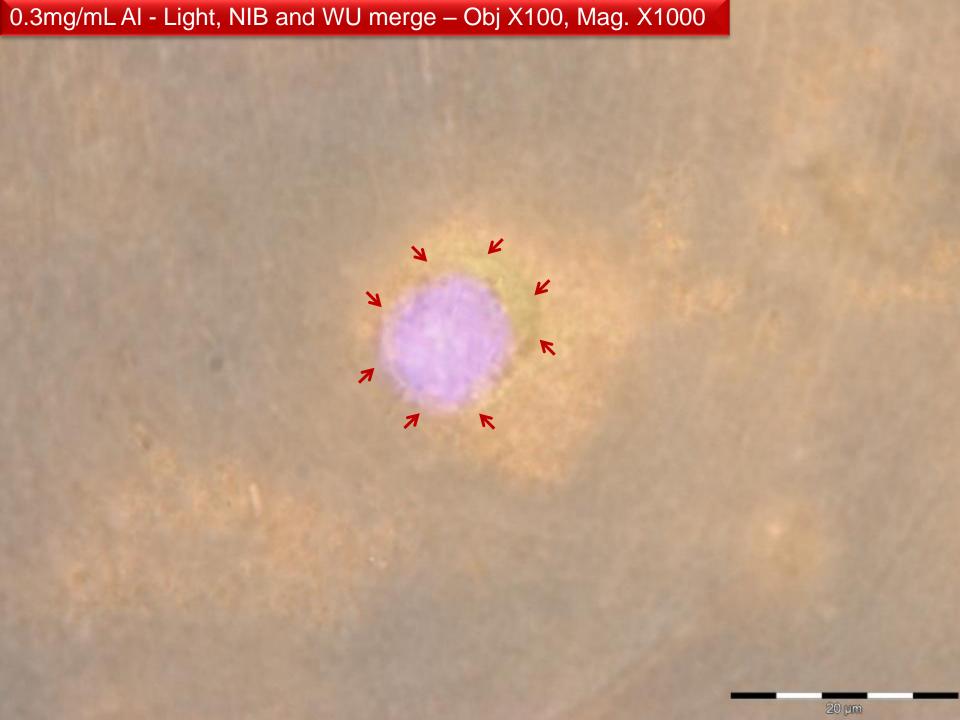


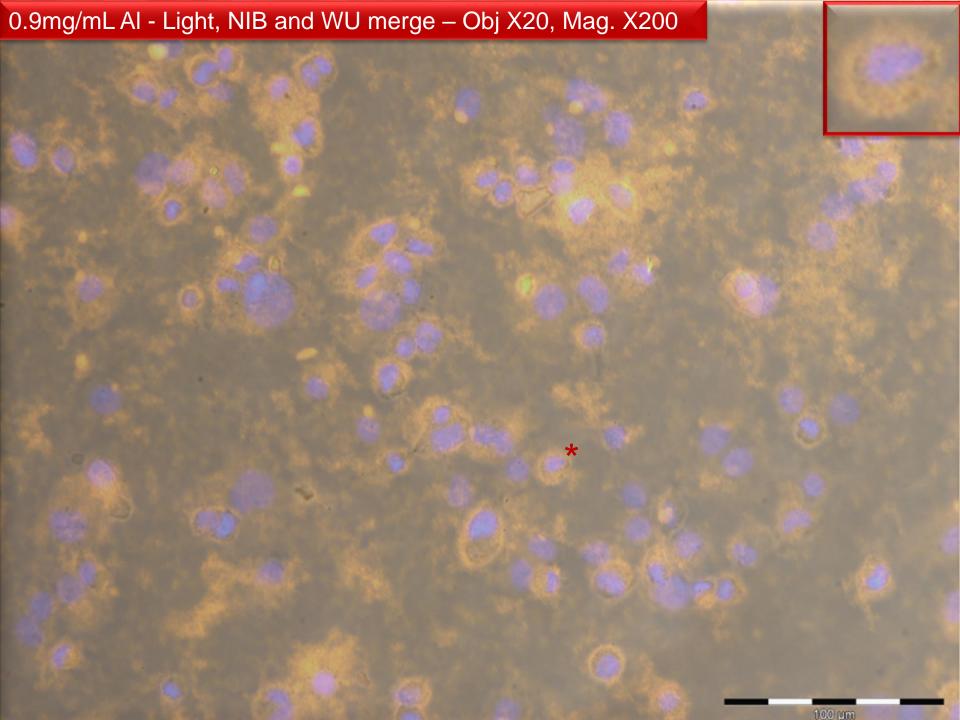
- Abundance of particles <0.22 μm remained consistent when concentration of Al increased.
- Significant differences observed between:
  - ❖ 0.5mg/mL & 0.9mg/mL (0.001 mg/mL vs. 0.0002 mg/mL, P=0.004)

Concentration of AI (mg/mL)	0.3	0.4	0.5	0.7	0.9	
	Recovery of AI (%)					
1-0.22μm	0.630	0.454	1.051	0.241	0.177	
<0.22µm	0.375	0.154	0.302	0.149	0.052	

Fig 11: The relative abundance of aluminium (%) within specific particle size fractions for Alhydrogel only vaccines containing an initial concentration of 0.3-0.9mg/mL Al following 1 hr incubation within R10 medium (37°C). Blue & purple boxes represent the % Al in the size fractions 1-0.22µm & >0.22µm respectively.

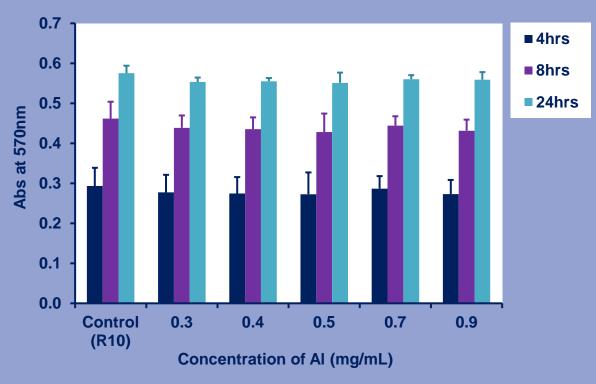








## Presto blue- Viability of macrophages exposed to various concentrations of AI in simulated vaccines.



- Null significance between treatment values and control at all time points.
- Cell viability was unaffected by all exposure regimes post 24hrs.

Concentration of Al (mg/mL)	0.3	0.4	0.5	0.7	0.9
Time	% Control				
4hrs	94.6	93.6	93.0	97.8	93.2
8hrs	95.0	94.3	92.8	96.2	93.5
24hrs	96.2	96.5	95.8	97.4	97.2

Fig 12: The viability of macrophages exposed to various concentrations of AI over a total duration of 24hrs. Error bars represent the ±SD of the measurement where n=5.

### **Conclusions**

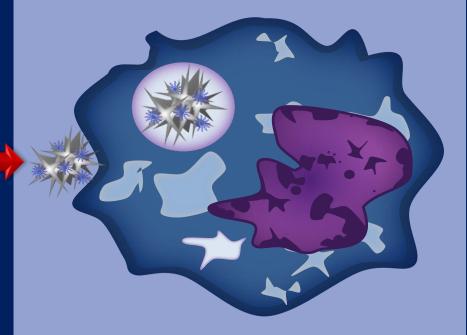


#### **Uptake**

> Uptake observed at low and high concentrations AI.

#### **Cell viability**

- ➤ Post uptake macrophage survival was not impaired following 24hrs incubation.
- > Translocation to lymph nodes highly likely at all concentrations studied.



#### Particle size in R10 (1-3µm)

- > 0.3mg/mL 87%
- > 0.9mg/mL 85%

#### References





















- [1] Baylor NW, Egan W, Richman P. Aluminium salts in vaccines US perspective. *Vaccine*, 2002, 29:S18-23.
- [2] Majgaard Jensen O, Koch C. On the effect of Al(OH)3 as an immunological adjuvant. APMIS,1988, 96(3):257-64.
- [3] Gupta RK, Rost BE, Relyveld E, Siber GR. Adjuvant properties of aluminium and calcium compounds. In: Powell, M.F. Newman, M.J. (eds.) Vaccine Design: The Subunit and Adjuvant Approach. 1995 . New York, US: Plenum.





















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